202. Conformationally Restricted Analogs of Platelet-Activating Factor (PAF)

by Paul Hadváry and Thomas Weller*

Pharmaceutical Research Department, F. Hoffmann-La Roche & Co., Ltd., CH-4002 Basle

(9.IX.86)

The synthesis of the five-membered cyclic phosphorodiamidic-acid derivatives 10 and 11 as well as the preparation of the six-membered cyclic phosphates 18, 19, 22–25, and phosphoramidates 27–32 is described. The effects of these conformationally restricted platelet-activating factor analogs on rabbit platelet aggregation are briefly discussed. The 2-oxo-1,3,2-dioxaphosphorinanes 19, 25, and 30 were found to be equally potent platelet-activating factor antagonists as the known thiazolium salt 33.

Introduction. – Platelet-activating factor (PAF, 1) is an ether phospholipid produced by a variety of different cell types [1] [2]. Recent studies have revealed that PAF is a potent platelet-activating, chemotactic, and hypotensive agent [3]; it also induces bronchoconstriction, increased vascular permeability, and gastric ulceration [4]. In general, PAF is believed to play an important role as mediator in anaphylactic and inflammatory processes. A PAF-specific cell surface receptor has been characterized recently [5]. Considerable efforts have been undertaken in order to i get information about structure-activity relationships [6–10], ii discover compounds with desired, selective physiological effects only, e.g. antihypertensive activity [6] [11], iii find inhibitors of the biosynthesis of PAF [12] [13], or iv identify antagonists of PAF [6] [14–25].



Here, we describe the synthesis and PAF-antagonistic properties of the conformationally restricted analogs 3–5 of PAF.

Syntheses. – Five-Membered Cyclic Analogs of Type 3. Because of the intrinsic instability of five-membered cyclic esters of phosphoric acid [26], e.g. 2, towards hydrolysis, we decided to prepare a cyclic phosphorodiamidic-acid derivative of type 3. Reaction of the (S)-amino alcohol 6¹) (Scheme 1) with N-(2-bromoethyl)phosphoramidic dichloride

¹) The amino alcohol **6** was prepared from (S)-1-O-octadecyl-glycerol [27] in three steps: *i*) TsCl, pyridine; *ii*) NaN₃, DMF; *iii*) H₂, Pd/C, THF. We thank Dr. R. Barner and Mr. G. Hirth for providing us with the experimental details of this reaction sequence [28].



 $(7)^2$) in the presence of Et₃N led to the formation of the two diastereoisomers 8 (27%) and 9 (26%), which could be easily separated by chromatography. Assignment of configuration at the P-atom is based on the observation that a P=O bond exhibits a deshielding effect on the protons in a 1,3-*cis*-relationship [30] as indicated in formula 9. Thus, the ¹H-NMR signal for H-C(5) in 9 appears at 4.65 ppm, while the corresponding proton in 8 resonates at 4.53 ppm.

Introduction of a positively charged N-function could be achieved by conversion of each diastereoisomer 8 and 9, in low yield, to the hygroscopic pyridinium salts 10 and 11, respectively, which turned out to be quite difficult to isolate. The ³¹ P-NMR spectrum of 11 showed the presence of a small amount of the diastereoisomer 10, which could not be removed by recrystallization.

Six-membered Cyclic Analogs of Type 4 and 5. The enantiomerically pure diol 14, keyintermediate in the synthesis of the six-membered cyclic PAF-analogs of types 4 and 5, was synthesized as shown in Scheme 2. Reaction of the benzylidene acetal 12 (prepared from (S)-1,2,4-butanetriol [31]) with octadecyl mesylate in the presence of solid KOH [32] produced the ether derivative 13. Removal of the protecting group to give 14 was accomplished by catalytic hydrogenation in AcOH. Treatment of the diol 14 with POCl₃ and Et₃N gave a single phosphochloridate 15. The known axial preference of the P-Cl bond in six-membered cyclic phosphates [33] [34] supports the structural assignment expressed in formula 15.

Exposure of 15 to 2-bromoethanol/Et₃N/4-(dimethylamino)pyridine (Me₂NPy) gave a 1:4 mixture of the two diastereoisomeric cyclic phosphates 16 and 17, separable by chromatography. The configuration of 16 and 17 was assigned using ³¹P-NMR spectroscopy, since it is well documented that the signals of 2-oxo-1,3,2-dioxaphosphorinanes with an axial P=O bond appear at lower field when compared to those of the isomers with an equatorial P=O bond [35]. Upon heating 16 in pyridine, the corresponding salt 18 was

²) This compound was available by heating 2-aminoethyl bromide hydrobromide in POCl₃ according to a protocol developed for the preparation of the corresponding 2-chloroethyl derivative [29].



produced in diastereoisomerically pure form. However, when 17 was subjected to the same reaction conditions, a 3:2 mixture of the pyridinium salts 19 and 18 was obtained, reflecting the axial preference of alkoxy substituents in six-membered cyclic phosphates under thermodynamic control [33] [34].

Because of the structural similarity of choline and 3-(trimethylammonio)phenol [36]³), the reaction of **15** with 3-(dimethylamino)phenol was briefly investigated (*Scheme 2*): in the presence of Et_3N/Me_2NPy , a 1:1 mixture⁴) of the two diastereoisomers **22** and **23** was formed, together with a small amount of the anhydride **26**. Chromatographic separation



was followed by methylation of 22 and 23, affording the quaternary ammonium salts 24 and 25. In the case of 23, the salt formation was again accompanied by isomerization yielding a 3:1 mixture of 25 and 24. The thermodynamically more stable isomer 22, on the other hand, could be converted to the single diastereoisomer 24.

To circumvent the isomerization problem just mentioned as well as to arrive at compounds of higher stability than the pyridinium salts 10, 11, 18, and 19, the phosphor-

³) All attempts to prepare the choline derivatives 20 and 21, starting either from 16 and 17 or directly from 15, failed.

⁴) In the absence of Me_2NPy , only 23 was observed, *i.e.* Me_2NPy accelerates the isomerization to the thermodynamically more stable diastereoisomer 22.



amidates 27–32 were prepared⁵). Treatment of 15, formed *in situ* from diol 14, with either 2-, 3-, or 4-(aminomethyl)pyridine in the presence of Et_3N/Me_2NPy proceeded with inversion at the P-atom leading almost exclusively to the equatorial intermediates 27–29. Methylation (CH₃I, 43°) then gave the corresponding pyridinium salts 30–32, whose ³¹P-NMR spectra revealed at most traces of the axial isomers.

Results. – To determine the PAF-antagonistic activity of 10, 11, 18, 19, 22–25, and 30–32, their ability to inhibit PAF-induced platelet aggregation in rabbit platelet-rich plasma (PRP) was examined. The same assay was used to measure eventual PAF-agonistic activity. The thiazolium salt 33 (CV-3988), shown to be a specific PAF-antagonist



by *Terashita et al.* [37], served as reference substance. From the results summarized in the *Table*, the following conclusions can be drawn:

i) With the exception of **22** and **32** at high concentrations, all compounds tested are devoid of proaggregatory activity.

ii) The five-membered cyclic phosphorodiamidic-acid derivatives 10 and 11 were found to be inactive as PAF-antagonists.

iii) In the series of the six-membered cyclic phosphates, the representatives with an equatorial side chain, *e.g.* **19** and **25**, exhibited PAF-antagonistic activity at significantly lower concentrations than the axial diastereoisomers **18** and **24**, respectively.

iv) A positively charged functional group seems to be a characteristic feature of an antagonist in this series, since the uncharged members 22 and 23 did not block PAF-induced platelet aggregation.

v) Comparison of the IC_{50} values of **30–32** demonstrates that PAF-antagonistic potency can be enhanced by increasing the distance between the P-atom and the positively charged, quaternary N-function.

vi) The conformationally restricted analogs 19, 25, and 30 were shown to be equipotent to the open-chain PAF-antagonist 33 (CV-3988).

⁵) In contrast to alkoxy groups, the alkylamino substituents show a strong preference for the equatorial position in 2-oxo-1,3,2-dioxaphosphorinanes [33] [34].

Compound	Inhibition of PAF ^a) IC ₅₀ [μM] ^b)	Proaggregation	
		Highest dose [µм]	Effect ^e)
10	>> 30	30	
11	> 10	10	_
18	17	30	
19 ^d)	2.1	30	-
22	≫ 30	30	aggregation followed by desaggregation
23	≫ 30	30	-
24	30	30	-
25 °)	3.2	30	-
30	4.0	30	-
31	6.5	30	-
32	12.0	30	shape change of the platelets
33	3.2	30	•

Table. Effects of Conformationally Restr	icted PAF Analogs on Platelet Aggregation		
in Rabbit Platelet-Rich Plasma (PRP)			

^a) Platelet aggregation was induced by 4 nм of PAF [38].

b) Determined from dose-response curves: Usually, PRP was preincubated with the test compound for 2 min at 37°.

^c) No effect observed: is indicated by a dash.

d) A 3:2 mixture of 19 and 18 was used.

e) A 3:1 mixture of 25 and 24 was used.

The skillful technical assistance of Mr. D. Cousot, Miss V. Schmid, Mr. D. Sprenger, and Mr. F. Gruber is gratefully acknowledged. We also thank our colleagues from the Central Research Department for determination of physical and analytical data: Dr. W. Arnold (NMR), Dr. L. Chopard (IR), Dr. A. Dirscherl (microanalyses), Dr. G. Englert (NMR), Dr. M. Grosjean (IR), Mr. W. Meister (MS), and Dr. M. Vecchi (GC).

Experimental Part

General. Reagent grade solvents (*Fluka, Merck*) were dried by passing through alumina *Woelm B-Super I*. POCl₃ (*Fluka*) was destilled before use. Evaporation means removal of solvent by use of a *Büchi* rotary evaporator at 40–50 °C/*in vacuo*. High vacuum (h.v.): 10^{-2} Torr. TLC: TLC plates coated with silica gel 60 *F*₂₅₄ (*Merck*); detection by UV (254 nm), by I₂ vapor, or by spraying with 50% H₂SO₄/EtOH followed by heating or with *Zinzadze* reagent [39] (for P-containing compounds). Flash chromatography (FC) [40]: silica gel 60 (*Merck*, 230–400 mesh ASTM). Medium pressure liquid chromatography (MPLC): *Lobar* columns, *LiChroprep Si 60* (40–63 µm, *Merck*), 1–4.5 bar (*CfG Pro Minent* pump). GC: *Varian 3700, SE 54* column (20 m). M.p.: uncorrected; *Büchi 510.* [α]²⁰₁₀: *Perkin Elmer 241* polarimeter, *c* in g/100 ml. IR spectra: *Nicolet-7199-FT-IR* spectrophotometer; in cm^{-1.} ¹H-NMR, ¹³C-NMR, and ³¹P-NMR spectra: *Bruker AS-250* (¹H(250 MHz)), *Bruker HX-270* (¹H(270 MHz), ¹³C(62.9 MHz)), or *Bruker WM-400* (¹H(400 MHz), ³¹P(162.0 MHz), ¹³C(100.62 MHz)); CDCl₃ solns. unless otherwise specified; δ values in ppm relative to tetramethylsilane (¹H- and ¹³C-NMR) as internal standard or H₃PO₄(³¹P-NMR) as external reference; coupling constants (*J*) in Hz. MS: *MS* 9 updated with a *Finnigan ZAB* console, data system *SS 200, VG Alirincham* (EI: 70 eV); *MM 7070 F*, data system 2050, *VG Alirincham* (CI: NH₃); *MS 902*, fast-atom gun *Kratos*, data system 2050, *VG Alirincham* (FAB, Xe-atom 6 keV, thioglycerol matrix (*Fluka*)); *m*/z (intensity in % of base peak (100%)).

(2R,5S)- and (2S,5S)-2-(2-Bromoethylamino)-5-(octadecyloxymethyl)-1,3,2-oxazaphospholidin-2-ones (8 and 9, resp.). To a suspension of 6¹) (576 mg, 1.68 mmol) in abs. THF (17 ml) under Ar was added at r.t. abs. Et₃N (492 µl) followed by 7²) (404 mg, 1.68 mmol). After 4 h at r.t., the white precipitates were removed by filtration through a sintered glass funnel. Evaporation of the filtrate afforded a solid crude product (725 mg), which was purified by MPLC (CH₂Cl₂/MeOH 49:1→29:1) to give 8 (233 mg, 27%) and 9 (223 mg, 26%).

8: M.p. 98–100° (AcOEt), $R_{\rm f}$ (CHCl₃/MeOH 9:1) 0.55, $[\alpha]_{20}^{20} = -5.9°$ (c = 0.51, THF). IR (KBr): 3226*m* (br.), 2919*s*, 2849*s*, 1468*m*, 1201*m*, 1123*m*, 1102*m*, 1078*m*, 720*w*. ¹H-NMR (400 MHz): 4.53 (*m*, 1 H); 3.67 (*dd*, J = 10.5, 6, 1 H); 3.55 (*dd*, J = 10.5, 5.5, 1 H); 3.51–3.31 (*m*, 9 H); 2.95 (*m*, 1 H); 1.56 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, J = 7, 3 H). ³¹P-NMR: +28.15. MS: 512 (5, M^{++} with ⁸¹Br), 510 (5, M^{++} with ⁷⁹Br), 431 (8), 417 (8), 244 (30), 242 (31), 43 (100). Anal. calc. for C₂₃H₄₈BrN₂O₃P (511.53): C 54.01, H 9.46, Br 15.62, N 5.48; found: C 54.35, H 9.71, Br 15.56, N 5.28.

9: M.p. 106–108°, $R_{\rm f}$ (CHCl₃/MeOH 9:1) 0.50, $[\alpha]_D^{20} = -0.9^{\circ}$ (c = 0.7, THF). IR (KBr): 3273*m* (br.), 2917*s*, 2850*s*, 1471*m*, 1214*s*, 1125*s*, 717*w*. ¹H-NMR (400 MHz): 4.65 (*m*, 1 H); 3.67–3.32 (*m*, 11 H); 2.92 (*m*, 1 H); 1.58 (*m*, 2 H); 1.26 (*m*, 30 H); 0.88 (t, J = 7, 3 H). ³¹P-NMR: +29.09. MS: 512 (2, M^{++} with ⁸¹Br), 510 (2, M^{++} with ⁷⁹Br), 431 (18), 417 (56), 43 (100).

I-{2-*[*(2R,5S)-5-(Octadecyloxymethyl)-2-oxo-1,3,2-oxazaphospholidin-2-yl)amino]ethyl}pyridinium Bromide (10). A soln. of 8 (100 mg, 0.19 mmol) in pyridine (2 ml) was kept under Ar at 45°. After 30 h, the solvents were azeotropically evaporated with cyclohexane to give a tan solide product (82 mg), which was successively washed with hexane, ACOEt, and acetone. The remaining material was then dissolved in CH₂Cl₂, filtered, and the filtrate evaporated to give, after drying under h.v., **10** (19 mg, 16%) as a tan, hygroscopic powder. M.p. 145° (dec.), $R_{\rm f}$ (CHCl₃/MeOH/H₂O 60:35:5) 0.45. IR (KBr): 3248m, 2918s, 2849s, 1634m, 1488m, 1221m, 1126m, 1070m. ¹H-NMR (270 MHz): 9.43 (m, 2 H); 8.47 (m, 1 H); 8.13 (m, 2 H); 5.29 (m, 1 H); 4.98 (m, 2 H); 4.40 (m, 2 H); 3.76–3.0 (m, 8 H); 1.52 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ³¹P-NMR: +29.63. FAB-MS: 510 (M^+ , cation).

 $l-\{2-[((2S,5S)-5-(Octadecyloxymethyl)-2-oxo-1,3,2-oxazaphospholidin-2-yl)amino]ethyl\}pyridinium Bro$ mide (11). Treatment of 9 (227 mg, 0.44 mmol) in pyridine (5 ml) according to the procedure described above $afforded 11 (88 mg, 33%) contaminated with traces of 10. M.p. 120–125° (dec.), <math>R_{\rm f}$ (CHCl₃/MeOH/H₂O 60:35:5) 0.3. IR (KBr): 3238m, 2918s, 2849s, 1635m, 1488m, 1468m, 1219m, 1128m, 1069m. ¹H-NMR (400 MHz): 9.54–9.40 (m, 2 H); 8.49 (m, 1 H); 8.12 (m, 2 H); 5.23 (m, 1 H); 5.0–4.8 (m, 2 H); 4.6–4.3 (m, 2 H); 3.65–2.9 (m, 8 H); 1.52 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ³¹P-NMR: +29.48, +29.64 (small peak). FAB-MS: 510 (M^{++} , cation).

(2S,4S)-4-(Octadecyloxymethyl)-2-phenyl-1,3-dioxane (13). A soln. of 12 (8.2 g, 42.2 mmol) in xylene (130 ml) was heated to reflux in the presence of powdered KOH (4.74 g, 84.4 mmol). During 2 h, the H₂O was collected in a *Dean-Stark* trap. After cooling to 90°, a soln. of octadecyl methanesulfonate (15.0 g, 43.1 mmol) in xylene (90 ml) was added dropwise. The resulting mixture was stirred at 90° for 46 h, then cooled to r.t., partitioned between Et₂O/AcOEt 2:1 and 10% aq. NaCl soln. and extracted (4 × 100 ml). The combined org. extracts were washed with 10% aq. NaCl soln., dried over Na₂SO₄, and evaporated. Purification by FC (250 g SiO₂, hexane/AcOEt 9:1) gave 13 (12.8 g, 68%) as a white solid. M.p. 58–59°, R_f (hexane/AcOEt 4:1) 0.56, $[a]_D^{20} = +0.67^\circ$ (c = 0.6, CHCl₃). IR (KBr): 2919s, 2849s, 1485w, 1125s, 1108s, 757m, 695m. ¹H-NMR (80 MHz): 7.55–7.2 (m, 5 H); 5.50 (br. s, 1 H); 4.3–3.3 (m, 7 H); 1.95–1.20 (m, 34 H); 0.88 (t, J = 6, 3 H). MS: 446 (24, M^{++}), 253 (10), 163 (100). Anal. calc. for C₂₉H₅₀O₃ (446.72): C 77.97, H 11.28; found: C 77.90, H 11.55.

(S)-4-(Octadecyloxy)butane-1,3-diol (14). A suspension of 13 (12.8 g, 28.7 mmol) in AcOH (560 ml) was warmed to 50° for 20 min. To the resulting soln. was added 10% Pd/C (0.6 g). Hydrogenation at 50° and normal pressure was complete within 20 min (1270 ml H₂). The catalyst was removed by filtration and the soln. evaporated. The residue was taken up in toluene (2 × 100 ml) and concentrated again to give 10.7 g of a colourless oil, which crystallized rapidly on standing. Recrystallization (AcOEt) afforded 14 (8.0 g, 78%). M.p. 60–61°, R_f (hexane/AcOEt) 0.11, $[\alpha]_D^{20} = +1.3$ (c = 0.15, CHCl₃). IR (KBr): 3410s, 2913s, 2848s, 1463m, 1125s, 1100m. ¹H-NMR (80 MHz): 3.82 (m, 3 H); 3.42 (m, 4 H); 2.25 (br. s, 2 H); 1.88–1.50 (m, 4 H); 1.25 (m, 30 H); 0.85 (t, J = 6, 3 H). MS: 358 (0, M^{++}), 283 (18), 252 (5), 75 (100). Anal. calc. for C₂₂H₄₆O₃ (358.61): C 73.69, H 12.93; found: C 73.63, H 12.78.

(2R,4S)- and (2S,4S)-2-(2-Bromoethoxy)-4-(octadecyloxymethyl)-1,3,2-dioxaphosphorinane 2-Oxides (16 and 17, resp.): General Procedure 1 (GP 1). To a soln. of 14 (1.80 g, 5 mmol) and Et₃N (1.46 ml, 10.5 mmol) in dry toluene (50 ml) was added POCl₃ (0.48 ml, 5.25 mmol). After stirring for 45 h at 50° under Ar, TLC showed a single new spot of 15. The precipitates were removed by filtration under Ar. To the filtrate was added Et₃N (0.93 ml, 6.67 mmol), 2-bromoethanol (0.49 ml, 7 mmol), and Me₂NPy (6 mg). The resulting mixture was stirred at 50° under Ar for 48 h. The precipitates were filtered off, the filtrate evaporated, and the residue purified by chromatography (hexane/AcOEt 1:1) to give 16 (0.21 g, 8%) and 17 (0.93 g, 35%).

16: M.p. 43°, R_f (hexane/AcOEt 1:1) 0.36. IR (KBr): 2918s, 2849s, 1299m, 1127m, 1065m, 720w. ¹H-NMR (400 MHz): 4.65 (m, 1 H); 4.49–4.33 (m, 4 H); 3.61 (m, 1 H); 3.59 (t, J = 6, 2 H); 3.51 (m, 1 H); 3.48 (t, J = 6.5, 2 H); 2.09 (m, 1 H); 1.86 (m, 1 H); 1.56 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ¹³C-NMR (100.6 MHz): 79.1 (d, J(P,C) = 7); 72.6 (t, J(P,C) = 11.7); 72.1 (t); 68.1 (t, J(P,C) = 7); 66.4 (t, J(P,C) = 5.3); 31.9 (t); 30.2 (t); 30.1 (t);

29.7 (*t*); 29.6 (*t*); 29.5 (*t*); 29.4 (*t*); 29.3 (*t*); 28.6 (*t*); 26.0 (*t*); 22.7 (*t*); 14.1 (*q*). ³¹P-NMR: -7.41. MS: 529 (1, $M^{++} + 1$ with ⁸¹Br), 527 (1, $M^{++} + 1$ with ⁷⁹Br), 447 (3), 277 (8), 275 (8), 54 (100).

17: M.p. 69° (hexane), R_{f} (hexane/AcOEt 1:1) 0.16. IR (KBr): 2918s, 2850s, 1272s, 1121m, 1069s, 1018s, 968s, 723w. ¹H-NMR (270 MHz): 4.73 (m, 1 H); 4.52–4.32 (m, 4 H); 3.61 (m, 2 H); 3.54 (t, J = 6.5, 2 H); 3.48 (t, J = 6.5, 2 H); 2.16 (m, 1 H); 2.03 (m, 1 H); 1.56 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ³¹P-NMR: -5.66. MS: 529 (<1, M^{++} + 1 with ⁸¹Br), 527 (<1, M^{++} + 1 with ⁷⁹Br), 447 (4), 277 (10), 275 (10), 71 (100). Anal. calc. for C₂₄H₄₈BrO₅P (527.52) · 0.1 C₆H₁₄: C 55.11, H 9.29, Br 14.90; found: C 55.33, H 9.06, Br 15.07.

The intermediate (2 R, 4 S)-2-chloro-4-(octadecyloxymethyl)-1,3,2-dioxaphosphorinane 2-oxide (15) might be isolated as a pale yellow foam upon simple concentration of a small part of the filtrate before addition of 2-bromoethanol. IR (KBr): 1301m, 1142m, 994m, 720w. ¹H-NMR (270 MHz): 4.68 (m, 1 H); 4.63–4.45 (m, 2 H); 3.65 (ddd, J = 11, 5, 2, 1 H); 3.55 (ddd, J = 11, 5.5, 2, 1 H); 3.48 (t, J = 7, 2 H); 2.31–1.88 (m, 2 H); 1.55 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 6.5, 3 H). MS: 440 (1, M^+ with ³⁷Cl), 438 (2, M^+ with ³⁵Cl), 189 (22), 187 (63), 71 (100).

(2S,4S)- and (2R,4S)-2-[3-(Dimethylamino)-phenoxy]-4-(octadecyloxymethyl)-1,3,2-dioxaphosphorinane 2-Oxides (22 and 23, resp.). The conversion of 14 (1.80 g, 5 mmol), POCl₃ (0.48 ml, 5.25 mmol), Et₃N (total amount 2.3 ml, 16.5 mmol), 3-(dimethylamino)phenol (686 mg, 5 mmol) and Me₂NPy (6 mg) in dry toluene (50 ml, then 75 ml) according to *GP 1* afforded, after chromatography with hexane/AcOEt 1:1, 22 (811 mg, 30%) and 23 (945 mg, 35%) as well as 26 (107 mg, 5%).

22: M.p. 63–64° (hexane), R_f (hexane/AcOEt 1:1) 0.43, $[\alpha]_{D}^{20} = -8.4°$ (c = 0.5, CHCl₃). IR (KBr): 2805w, 1606s, 1574m, 1503s, 1291s, 1145s, 997s, 986s. ¹H-NMR (400 MHz): 7.16 (m, 1 H); 6.59 (m, 2 H); 6.51 (m, 1 H); 4.70 (m, 1 H); 4.53–4.40 (m, 2 H); 3.64 (ddd, J = 10.5, 5, 1.5, 1 H); 3.52 (ddd, J = 10.5, 5.5, 2, 1 H); 3.47 (t, J = 6.5, 2 H); 2.94 (br. s, 6 H); 2.15 (m, 1 H); 1.91 (m, 1 H); 1.55 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ¹³C-NMR: 151.9 (s); 151.5 (s, J(P,C) = 6.4); 129.9 (d); 109.0 (d); 106.8 (d, J(P,C) = 4.9); 103.4 (d, J(P,C) = 5.8); 79.4 (d, J(P,C) = 7.2); 72.7 (t, J(P,C) = 9.4); 72.0 (t); 68.3 (t, J(P,C) = 7.2); 40.3 (q); 31.9 (t); 29.7 (t); 29.6 (t); 29.5 (t); 29.4 (t); 28.6 (t); 28.5 (t); 26.1 (t); 22.7 (t); 14.1 (q). ³¹P-NMR: -12.95. MS: 539 (100, M^+). Anal. calc. for C₃₀H₅₄NO₅P (539.74): C 66.76, H 10.08, N 2.60; found: C 66.77, H 10.02, N 2.61.

23: M.p. 57-58° (hexane), $R_{\rm f}$ (hexane/AcOEt 1:1) 0.18, $[\alpha]_{D}^{20} = +13.0^{\circ}$ (c = 1, CHCl₃). IR (KBr): 2817w, 1606s, 1572m, 1507m, 1294s, 1073s, 998s. ¹H-NMR (400 MHz): 7.14 (m, 1 H); 6.57-6.48 (m, 3 H); 4.77 (m, 1 H); 4.57-4.43 (m, 2 H); 3.64 (ddd, J = 10.5, 5, 2, 1 H); 3.57 (ddd, J = 10.5, 6.5, 1, 1 H); 3.44 (t, J = 7, 2 H); 2.94 (br. s, 6 H); 2.18–2.02 (m, 2 H); 1.53 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ¹³C-NMR (100.6 MHz): 151.8 (s); 151.7 (s, J(P,C) = 7.1); 129.7 (d); 109.2 (d); 107.5 (d, J(P,C) = 4); 104.1 (d, J(P,C) = 5.5); 78.3 (d, J(P,C) = 6.9); 71.9 (t); 71.8 (t, J(P,C) = 6.7); 66.9 (t, J(P,C) = 8.2); 40.4 (q); 31.9 (t); 29.7 (t); 29.5 (t); 29.4 (t); 27.7 (t); 27.6 (t); 26.1 (t); 22.7 (t); 14.1 (q). ³¹P-NMR: -11.31. MS: 539 (100, M^{+*}). Anal. calc. for C₃₀H₅₄NO₅P (539.74): C 66.76, H 10.08, N 2.60; found: C 66.74, H 10.16, N 2.60.

2.2'-Oxybis[4-(octadecyloxymethyl)-1,3,2-dioxaphosphorinane 2-Oxide] (26). M.p. 87-89° (Et₂O), R_f (hexane/AcOEt 1:1) 0.25. IR (KBr): 1313s, 1302s, 1142m, 1067s, 976s. ¹H-NMR (270 MHz): 4.88 (m, 2 H); 4.75 (m, 2 H); 4.52 (m, 2 H); 3.67-3.42 (m, 8 H); 2.20 (m, 2 H); 1.90 (m, 2 H); 1.55 (m, 4 H); 1.25 (m, 60 H); 0.88 (t, J = 7, 6 H). FAB-MS: 823 (M^{++} + 1).

(2R,4S)-4-(Octadecyloxymethyl)-2-[(4-pyridylmethyl)amino]-1,3,2-dioxaphosphorinane 2-Oxide (27). Treatment of 14 (7.0 g, 19.5 mmol) in toluene (2 × 200 ml) with POCl₃ (1.9 ml, 21 mmol), Et₃N (5.9 and 2.9 ml, 63 mmol), and 4-(aminomethyl)pyridine (2 ml, 20 mmol) according to *GP* 1 gave, after FC (CH₂Cl₂/MeOH 19:1), 27 (5.65 g, 56%). M.p. 70–72° (hexane), R_{f} (CH₂Cl₂/MeOH 9:1) 0.46. $[\alpha]_{D}^{20} = -3.8°$ (c = 1, CHCl₃). IR (KBr): 3196m, 1604m, 1563m, 1232s, 1126m, 1081s. ¹H-NMR (400 MHz): 8.55 (m, 2 H); 7.28 (m, 2 H); 4.75 (m, 1 H); 4.57 (m, 1 H); 4.32 (m, 1 H); 4.28 (m, 2 H); 3.54 (ddd, J = 10.5, 4.5, 2, 1 H); 3.48 (ddd, J = 10.5, 5, 1, 1 H); 3.43 (t, J = 6.5, 2 H); 3.40 (m, 1 H); 2.05 (m, 1 H); 1.80 (m, 1 H); 1.52 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ³¹P-NMR: +5.93, +3.24 (very small peak). FAB-MS: 511 (M^{++} +1). Anal. calc. for C₂₈H₅₁N₂O₄P (510.70): C 65.85, H 10.07, N 5.49, P 6.07; found: C 65.58, H 10.14, N 5.32, P 5.91.

(2R,4S)-4-(Octadecyloxymethyl)-2-[(3-pyridylmethyl)amino]-1,3,2-dioxaphosphorinane 2-Oxide (28). Conversion of 14 (717 mg, 2 mmol), POCl₃ (0.19 ml, 2.1 mmol), Et₃N (0.59 and 0.29 ml, 6.3 mmol), and 3-(aminomethyl)pyridine (0.20 ml, 2 mmol) in toluene (total amount 35 ml) according to *GP 1* yielded, after MPLC (CH₂Cl₂/MeOH 10:1), 28 (709 mg, 69%). M.p. 60–62°, R_f (CH₂Cl₂/MeOH 9:1) 0.48. IR (KBr): 3212*m*, 1577*w*, 1242*s*, 1077*s*, 1055*s*. ¹H-NMR (270 MHz): 8.56 (*m*, 1 H); 8.52 (*m*, 1 H); 7.75 (*m*, 1 H); 7.28 (*m*, 1 H); 4.73 (*m*, 1 H); 4.56 (*m*, 1 H); 4.30 (*m*, 1 H); 4.18 (*m*, 2 H); 3.56 (ddd, J = 10.5, 4.5, 2, 1 H); 3.48 (*m*, 1 H); 3.44 (*t*, J = 7, 2 H); 3.42 (*m*, 1 H); 2.02 (*m*, 1 H); 1.81 (*m*, 1 H); 1.53 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, J = 7, 3 H). ¹³C-NMR (100.6 MHz): 148.9 (*d*); 148.8 (*d*); 135.2 (*d*); 134.9 (*s*); 123.4 (*d*); 76.4 (*d*, J(P,C) = 5); 72.8 (*t*, J(P,C) = 10); 71.9 (*t*); 65.9 (*t*, J(P,C) = 6); 41.9 (*t*); 31.9 (*t*); 29.7 (*t*); 29.6 (*t*); 29.5 (*t*); 29.4 (*t*); 28.6 (*t*); 28.5 (*t*); 26.1 (*t*); 22.7 (*t*); 14.1 (*q*).

³¹P-NMR: +5.87, +3.25 (very small peak). MS: 510 (64, M^+), 242 (80), 43 (100). Anal. calc. for C₂₈H₅₁N₂O₄P (510.70): C 65.85, H 10.07, N 5.49, P 6.07; found: C 65.92, H 10.62, N 5.47, P 6.18.

(2R,4S)-4-(*Octadecyloxymethyl*)-2-{(2-pyridylmethyl)amino}-1,3,2-dioxaphosphorinane 2-Oxide (29). Reaction of 14 (717 mg, 2 mmol) with POCl₃ (0.19 ml, 2.1 mmol), Et₃N (0.59 and 0.29 ml, 6.3 mmol), and 2-(aminomethyl)pyridine (0.20 ml, 2 mmol) in toluene (total amount 35 ml) according to *GP* 1 gave, after MPLC (CH₂Cl₂/MeOH 49:1), 29 (325 mg, 32%). M.p. 82–83°, R_f (CH₂Cl₂/MeOH 9:1) 0.14, $[\alpha]_{20}^{20}$ = +6.0° (*c* = 1, CHCl₃). IR (KBr): 3207*m*, 1591*w*, 1223*s*, 1122*s*, 1064*s*. ¹H-NMR (270 MHz): 8.54 (*m*, 1 H); 7.65 (*m*, 1 H); 7.36 (*m*, 1 H); 7.15 (*m*, 1 H); 4.74 (*m*, 1 H); 4.58 (*m*, 1 H); 4.38–4.23 (*m*, 3 H); 4.16 (*m*, 1 H); 3.57 (*ddd*, *J* = 10.5, 4.5, 2, 1 H); 3.48 (*m*, 1 H); 3.43 (*t*, *J* = 6.5, 2 H); 2.05 (*m*, 1 H); 1.82 (*m*, 1 H); 1.53 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, *J* = 7, 3 H). ³¹P-NMR: +6.04. MS: 510 (18, *M*⁺⁺), 242 (54), 70 (100). Anal. calc. for C₂₈H₅₁N₂O₄P (510.70): C 65.85, H 10.07, N 5.49, P 6.07; found: C 65.55, H 10.16, N 5.25, P 5.63.

I-{2-*[*(2R,4S)-4-(Octadecyloxymethyl)-2-oxo-1,3,2-dioxaphosphorinan-2-yl)oxy]ethyl}pyridinium Bromide (18). A soln. of 16 (150 mg, 0.28 mmol) in pyridine (3 ml) was stirred at 50° under Ar. After 26 h, the solvents were evaporated. The residue was washed with AcOEt (2×) and recrystallized from acetone (2×) to give 18 (63 mg, 37%). M.p. 75° (dec. > 85°), R_f (CHCl₃/MeOH/H₂O 60:35:5) 0.50. IR (KBr): 1634m, 1580m, 1292s, 995s, 965s. ¹H-NMR (270 MHz): 9.76 (m, 2 H); 8.48 (m, 1 H); 8.07 (m, 2 H); 5.55 (m, 2 H); 4.81–4.32 (m, 5 H); 3.62–3.42 (m, 4 H); 2.14 (m, 1 H); 1.85 (m, 1 H); 1.52 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). FAB-MS: 526 (M^{++} , cation). Anal. calc. for C₂₉H₅₃BrNO₅P (606.62): C 57.42, H 8.81, N 2.31; found: C 57.24, H 8.72, N 2.06.

Treatment of **17** (150 mg, 0.28 mmol) in pyridine/CH₃CN 1:1 (5 ml) with 4 Å-molecular sieves (0.5 g) in the same way as described above led to **19/18** 3:2 (35 mg, 20%). M.p. 25–30°. IR (KBr): 1634*m*, 1580*m*, 1289*s*, 965*s*. ¹H-NMR (270 MHz): 9.76, 9.62 (2*m*, ratio 2:3, 2 H); 8.50 (*m*, 1 H); 8.08 (*m*, 2 H); 5.60–5.38 (*m*, 2 H); 4.80–4.30 (*m*, 5 H); 3.63–3.41 (*m*, 4 H); 2.13 (*m*, 1 H); 1.90 (*m*, 1 H); 1.55 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, J = 7, 3 H). FAB-MS: 526 (M^{++} , cation).

Trimethyl {3-{((2S,4S)-4-(octadecyloxymethyl)-2-oxo-1,3,2-dioxaphosphorinan-2-yl)oxy}phenyl}ammonium *Iodide* (24): *General Procedure 2* (GP 2). A soln. of **22** (100 mg, 0.19 mmol) in CH₃I (5 ml) was kept at 50° unter Ar for 96 h. The solvents were then evaporated. The solid residue was washed with hot hexane and dried under h.v. to give **24** (115 mg, 91%). M.p. 110° (> 129°, dec.). IR (KBr): 1604*m*, 1490*m*, 1236*m*, 1224*m*, 968*s*. ¹H-NMR (400 MHz): 8.20 (br. *s*, 1 H); 7.79 (*m*, 1 H); 7.64 (*m*, 2 H); 5.10 (*m*, 1 H); 4.90 (*m*, 1 H); 4.53 (*m*, 1 H); 4.02 (br. *s*, 9 H); 3.63 (*m*, 2 H); 3.51 (*m*, 2 H); 2.24 (*m*, 1 H); 2.01 (*m*, 1 H); 1.55 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, *J* = 7, 3 H). ¹³C-NMR (100.6 MHz): 151.7 (*s*, *J*(P,C) = 5); 148.0 (*s*); 132.0 (*d*); 122.1 (*d*); 116.5 (*d*); 113.4 (*d*, *J*(P,C) = 7); 80.7 (*d*, *J*(P,C) = 70; 72.4 (*t*, *J*(P,C) = 10); 72.1 (*t*); 69.6 (*t*, *J*(P,C) = 7); 57.9 (*q*); 31.9 (*t*); 29.7 (*t*); 29.5 (*t*); 29.4 (*t*): 27.9 (*t*); 27.8 (*t*); 26.1 (*t*); 22.7 (*t*); 14.1 (*q*). FAB-MS: 554 (*M*⁺⁺, cation). Anal. calc. for C₃₁H₅₇INO₅P (681.67): C 54.62, H 8.43, I 18.62, N 2.05; found: C 54.25, H 8.66, I 18.74, N 2.09.

Treatment of **23** (100 mg, 0.19 mmol) in CH₃I (5 ml) according to *GP 2* led to a 3:1 mixture **25/24** (115 mg, 91%). M.p. 132–133°. ¹H-NMR (270 MHz): 8.20, 7.94 (2*m*, ratio 1:3, 1 H); 7.85, 7.79 (2*m*, ratio 3:1, 1 H); 7.63 (*m*, 1 H); 7.48 (*m*, 1 H); 5.10 (*m*, 0.25 H); 4.95–4.40 (*m*, 2.75 H); 4.02 (br. *s*, 9 H); 3.77–3.60 (*m*, 2 H); 3.47 (*m*, 2 H); 2.42–1.85 (*m*, 2 H); 1.55 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, J = 7, 3 H). FAB-MS: 554 (M^{++} , cation). Anal. calc. for C₃₁H₅₇INO₅P (681.67): C 54.62, H 8.43, I 18.62, N 2.05; found: C 54.69, H 8.54, I 18.33, N 1.97.

I-Methyl-4-{ $[((2' R,4' S)-4'-(octadecyloxymethyl)-2'-oxo-1',3',2'-dioxaphosphorinan-2'-yl)amino]methyl}$ pyridinium Iodide (**30**). Conversion of**27**(1.5 g, 2.94 mmol) in CH₃I (**30**ml) according to*GP 2*(reaction time 1 h) gave, after recrystallization from acetone,**30** $(1.65 g, 86%). M.p. 75° (dec.). IR (KBr): 3188m, 1643m, 1577w, 1519w, 1224m, 1069s. ¹H-NMR (270 MHz): 9.08 (m, 2 H); 8.09 (m, 2 H); 5.38 (m, 1 H); 4.75 (m, 1 H); 4.57 (br. s, 3 H); 4.50–4.28 (m, 4 H); 3.60 (dd, J = 10.5, 5, 1 H); 3.49 (m, 1 H); 3.46 (t, J = 6.5, 2 H); 2.14–1.78 (m, 3 H); 1.55 (m, 2 H); 1.25 (m, 30 H); 0.88 (t, J = 7, 3 H). ³¹P-NMR: +5.44, +3.08 (very small peak). FAB-MS: 525 (<math>M^{++}$, cation). Anal. calc. for C₂₉H₅₄IN₂O₄P (652.64): C 53.37, H 8.34, N 4.29, P 4.75; found: C 52.84, H 8.35, N 4.49, P 4.85.

I-Methyl-3-{ $f((2' R, 4' S) - 4' - (octade cyloxymethyl) - 2' - oxo-1', 3', 2' - dioxaphosphorinan-2' -yl)amino]methyl}-pyridinium Iodide ($ **31**). From**28**(511 mg, 1 mmol) and CH₃I (5 ml),**31**(580 mg, 89%) was obtained according to*GP 2* $(reaction time 90 min). M.p. 57° (acetone, dec.), <math>R_{f}(CH_{2}Cl_{2}/MeOH 9:1) 0.17, [\alpha]_{D}^{20} = +6° (c = 1, CHCl_{3}).$ IR (KBr): 3244*m*, 1636*m*, 1507*m*, 1241*s*, 1066*s*. ¹H-NMR (400 MHz): 9.23 (br. *s*, 1 H); 9.00 (*m*, 1 H); 8.55 (*m*, 1 H); 8.06 (*m*, 1 H); 5.42 (*m*, 1 H); 4.74 (*m*, 1 H); 4.54 (br. *s*, 3 H); 4.50–4.32 (*m*, 4 H); 3.61 (*m*, 1 H); 3.52 (*m*, 1 H); 3.47 (*t*, *J* = 7, 2 H); 2.10 (*m*, 1 H); 1.82 (*m*, 1 H); 1.54 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*, *J* = 7, 3 H). ¹³C-NMR (100.6 MHz): 144.7 (*d*); 144.5 (*d*); 143.7 (*d*); 141.6 (*s*); 127.8 (*d*); 76.8 (*d*, *J*(P,C) = 5); 72.9 (*t*, *J*(P,C) = 10); 71.9 (*t*); 66.4 (*t*); 49.4 (*q*); 41.7 (*t*); 31.9 (*t*); 29.7 (*t*); 29.5 (*t*); 29.4 (*t*); 26.1 (*t*); 22.7 (*t*); 14.1 (*q*). ³¹P-NMR : +5.42, +3.09 (very small peak). FAB-MS: 525 (*M*⁺⁺, cation). Anal. calc. for C₂₉H₅₄IN₂O₄P (652.64): C 53.37, H 8.34, I 19.44, N 4.29; found: C 53.26, H 8.69, I 19.15, N 4.15.

I-Methyl-2-{ $[((2' R, 4' S)-4'-(octadecyloxymethyl)-2'-oxo-1', 3', 2'-dioxaphosphorinan-2'-yl)amino]methyl}-pyridinium Iodide ($ **32**). Reaction of**29**(217 mg, 0.42 mmol) in CH₃I (4 ml) according to*GP*2 afforded**32**(153 mg, 55%). M.p. 58° (acetone, dec.). IR (KBr): 3181*m*, 1631*m*, 1514*w*, 1245*s*, 1067*s*. ¹H-NMR (270 MHz): 9.09 (*m*, 1 H); 8.46 (*m*, 1 H); 8.33 (*m*, 1 H); 7.95 (*m*, 1 H); 5.23 (*m*, 1 H); 4.75 (*m*, 3 H); 4.58-4.30 (*m*, 2 H); 4.50 (br.*s*, 3 H); 3.63 (*m*, 1 H); 3.50 (*m*, 1 H); 3.46 (*t*,*J*= 6.5, 2 H); 2.16 (*m*, 1 H); 1.88 (*m*, 1 H); 1.55 (*m*, 2 H); 1.25 (*m*, 30 H); 0.88 (*t*,*J*= 7, 3 H). ³¹P-NMR: +5.14, +2.54 (very small peak). FAB-MS: 525 (*M*⁺⁺, cation).

REFERENCES

- J. Benveniste, B. B. Vargaftig, in 'Ether Lipids, Biochemical and Biomedical Aspects', Eds. H. K. Mangold and F. Paltauf, Academic Press, New York, 1983, Chapt. 18, p. 355.
- [2] F. Snyder, Ann. Rep. Med. Chem. 1982, 17, 243 and ref. cit. therein.
- [3] F. Snyder, Med. Res. Rev. 1985, 5, 107.
- [4] A. Rosam, J. L. Wallace, B. J. R. Whittle, Nature (London) 1986, 319, 54; J. L. Wallace, B. J. R. Whittle, Br. J. Pharmacol. 1986, 87, 92 P.
- [5] S.-B. Hwang, C.-S.C. Lee, M.J. Cheah, T.Y. Shen, Biochemistry 1983, 22, 4756; G. Lambrecht, M.J. Parnham, Br. J. Pharmacol. 1986, 87, 287.
- [6] M.C. Venuti, Ann. Rep. Med. Chem. 1985, 20, 193 and ref. cit. therein.
- [7] G. Hirth, H. Saroka, W. Bannwarth, R. Barner, Helv. Chim. Acta 1983, 66, 1210.
- [8] H.-P. Kertscher, G. Ostermann, Pharmazie 1985, 40, 55.
- [9] H. Hayashi, I. Kudo, K. Inoue, K. Onozaki, S. Tsushima, H. Nomura, S. Nojima, J. Biochem. 1985, 97, 1737.
- [10] F. Heymans, M.-C. Borrel, C. Broquet, J. Lefort, J.-J. Godfroid, J. Med. Chem. 1985, 28, 1094.
- [11] A. Wissner, R. E. Schaub, P. E. Sum, C. A. Kohler, B. M. Goldstein, J. Med. Chem. 1986, 29, 328, 1315; ibid. 1985, 28, 1181.
- [12] K. M. Rupprecht, M. M. Ponpipom, J. C. Robbins, M. H. Lam, T. Y. Shen, Pharmacologist 1985, 27, 176.
- [13] J. C. Robbins, B. H. Ma Choy, M. H. Lam, M. M. Ponpipom, K. M. Rupprecht, T. Y. Shen, Fed. Proc. 1985, 44, 1269.
- [14] E. Kornecki, Y. H. Ehrlich, R. H. Lenox, Science 1984, 226, 1454.
- [15] T.Y. Shen, S.S. Yang, S.B. Hwang, to Merck & Co. Inc., 1985, Eur. Pat. Appl. 142801.
- [16] S.B. Hwang, M.H. Lam, T. Biftu, T.R. Beattie, T.Y. Shen, J. Biol. Chem. 1985, 260, 15639.
- [17] T. Miyamoto, H. Ohno, T. Yano, T. Okada, M. Hamanaka, A. Kawasaki, in 'Adv. Prostaglandin, Thromboxane, and Leukotriene Research', Eds. O. Hayaishi and S. Yamamoto, Raven Press, New York, 1985, Vol. 15, p. 719.
- [18] A. Tokumura, H. Homma, D.J. Hanahan, J. Biol. Chem. 1985, 260, 12710; D.B. Buxton, D.J. Hanahan, M.S. Olson, Biochem. Pharmacol. 1986, 35, 893.
- [19] M. L. Lee, C. M. Winslow, C. Jaeggi, F. D'Aries, G. Frisch, C. Farley, M. K. Melden, D. A. Handley, R. N. Saunders, *Prostaglandins* 1985, 30, 690; C. M. Winslow, S. R. Vallespir, G. E. Frisch, F. J. D'Aries, A. K. De Lillo, W. J. Houlihan, V. Parrino, G. Schmitt, R. N. Saunders, *ibid.* 1985, 30, 697; P. Sedivy, C. G. Caillard, A. Floch, F. Folliard, S. Mondot, C. Robaut, B. Terlain, *ibid.* 1985, 30, 688; P. Braquet, *ibid.* 1985, 30, 687; D. A. Handley, R. W. Deacon, J. C. Tomesch, J. M. Koletar, R. N. Saunders, *Fed. Proc.* 1986, 45, 685.
- [20] M. Steiner, R. Landolfi, N.C. Motola, J.G. Turcotte, Biochem. Biophys. Res. Commun. 1985, 133, 851.
- [21] M. Okamoto, K. Yoshida, I. Uchida, M. Kohsaka, H. Aoki, Chem. Pharm. Bull. 1986, 34, 345.
- [22] M. Okamoto, K. Yoshida, M. Nishikawa, T. Ando, M. Iwami, M. Kohsaka, H. Aoki, J. Antibiot. 1986, 39, 198.
- [23] K. Burri, R. Barner, J.-M. Cassal, P. Hadváry, G. Hirth, K. Müller, Prostaglandins 1985, 30, 691; P. Hadváry, H. R. Baumgartner, ibid. 1985, 30, 694.
- [24] T. Biftu, to Merck & Co. Inc., 1985, Eur. Pat. Appl. 154887.
- [25] H. Disselnkötter, F. Lieb, H. Oediger, D. Wendisch, Arch. Pharm. 1985, 318, 695.
- [26] F.H. Westheimer, Acc. Chem. Res. 1968, 1, 70.
- [27] G. Hirth, R. Barner, Helv. Chim. Acta. 1982, 65, 1059.
- [28] R. Barner, G. Hirth (F. Hoffmann-La Roche & Co. AG, Central Research Units), unpublished results.
- [29] B.S. Drach, A. D. Sinitsa, Zh. Obshch. Khim. 1968, 38, 1325; H.J. Roth, A.R. Lenig, W. Stock, Arch. Pharm. 1981, 314, 85.

- [30] D. B. Cooper, C. R. Hall, J. M. Harrison, T. D. Inch, J. Chem. Soc., Perkin Trans. 1 1977, 1969.
- [31] E. Hungerbühler, D. Seebach, D. Wasmuth, Angew. Chem., Int. Ed. 1979, 18, 958; T. Tsuri, S. Kamata, Tetrahedron Lett. 1985, 26, 5195.
- [32] W.J. Baumann, H.K. Mangold, J. Org. Chem. 1964, 29, 3055.
- [33] A.J. Kirby, 'The Anomeric Effect and Related Stereoelectronic Effects at Oxygen', Springer Verlag, Berlin, 1983, p. 26ff.
- [34] B. E. Maryanoff, R. O. Hutchins, C. A. Maryanoff, in 'Topics in Stereochemistry', Eds. N. L. Allinger and E. E. Eliel, J. Wiley and Sons, New York, 1979, Vol. 11, pp. 187–326.
- [35] W. Stec, M. Mikołajczyk, Tetrahedron 1973, 29, 539; W. Stec, A. Łopusiński, *ibid.* 1973, 29, 547; J. Engels,
 E.-J. Schlaeger, J. Med. Chem. 1977, 20, 907; J. A. Mosbo, J. G. Verkade, J. Org. Chem. 1977, 42, 1549; D.G.
 Gorenstein, R. Rowell, J. Am. Chem. Soc. 1979, 101, 4925; J. Baraniak, K. Lesiak, M. Sochacki, W.J. Stec,
 ibid. 1980, 102, 4533; D.G. Gorenstein, R. Rowell, J. Findley, *ibid.* 1980, 102, 5077; R.O. Day, D.G.
 Gorenstein, R. R. Holmes, Inorg. Chem. 1983, 22, 2192.
- [36] I. B. Wilson, C. Quan, Arch. Biochem. Biophys. 1958, 73, 131; G. H. Cocolas, J. G. Cranford, H. S. Yun Choi, J. Med. Chem. 1974, 17, 938; Y. Ashani, H. Leader, L. Raveh, R. Bruckstein, M. Spiegelstein, ibid. 1983, 26, 145.
- [37] Z. Terashita, S. Tsushima, Y. Yoshioka, H. Nomura, Y. Inada, K. Nishikawa, Life Sci. 1983, 32, 1975.
- [38] P. Hadváry, H. R. Baumgartner, Thromb. Res. 1983, 30, 143.
- [39] C. Zinzadze, Ind. Eng. Chem. 1935, 7, 227; J.C. Dittmer, R.L. Lester, J. Lipid Res. 1964, 5, 126.
- [40] W.C. Still, M. Kahn, A. Mitra, J. Org. Chem. 978, 43, 2923.